

ENVIRONMENTAL SOURCES OF NTM & REDUCING EXPOSURE

Rachel Thomson

Gallipoli Medical Research Institute, University of Queensland

Brisbane, Australia

DISCLOSURES

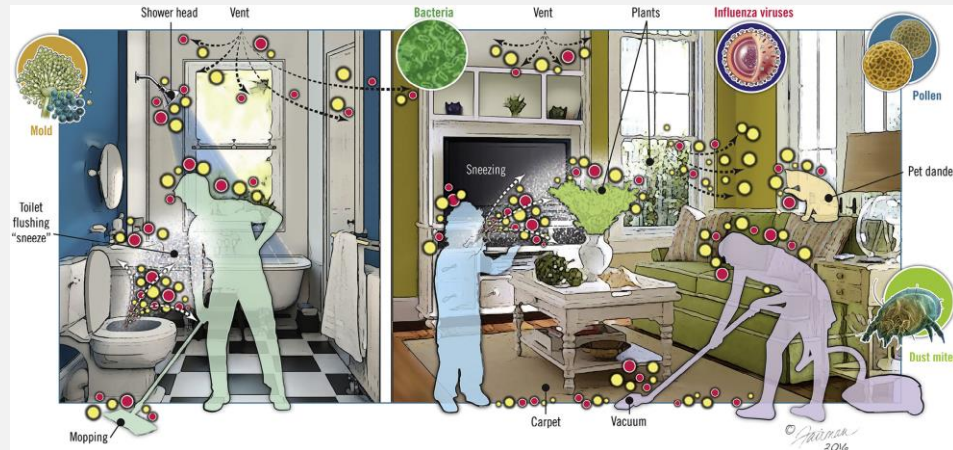
- Advisory boards- Insmmed, Savara
- Clinical trials – Insmmed, Aradigm, BMS, Savara, ResTORbio, Zambon
- CME talks - Menarini, GSK, AstraZeneca

INTRODUCTION

- Nontuberculous mycobacteria are ubiquitous in the environment.
- Household and hospital tap water, bathrooms, potting soil, and garden soil are infection sources identified by matching genotypic profiles of clinical and environmental isolates.
- Mycobacterial transmission routes include water, soil and cough aerosols

S110

M.K. Ijaz et al. / American Journal of Infection Control 44 (2016) S109-S120



WATER

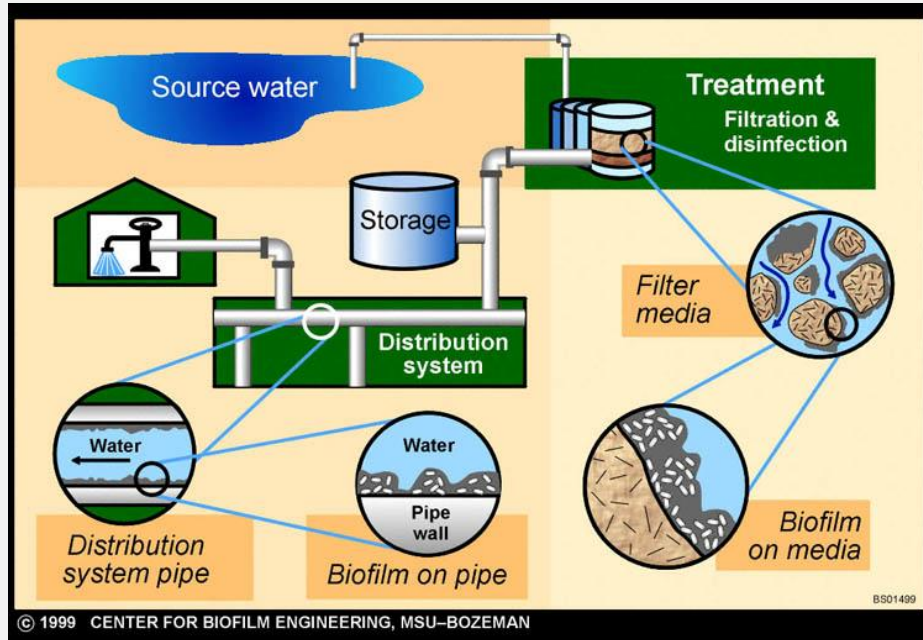
**IN WINE THERE IS TRUTH. IN
BEER THERE IS STRENGTH.**



**IN WATER THERE IS
BACTERIA.**

- Wide variety of NTM species identified in water samples – not all associated with disease
- Genotyping to match patient with water isolates
- Household/municipal water systems - *M. avium*, *M. abscessus*, *M. kansasii*, *M. lentiflavum*
- Hospital water - *M. porcinum*, *M. avium*, *M. abscessus*, *M. fortuitum*, *M. chimaera*
 - not *M. intracellulare*

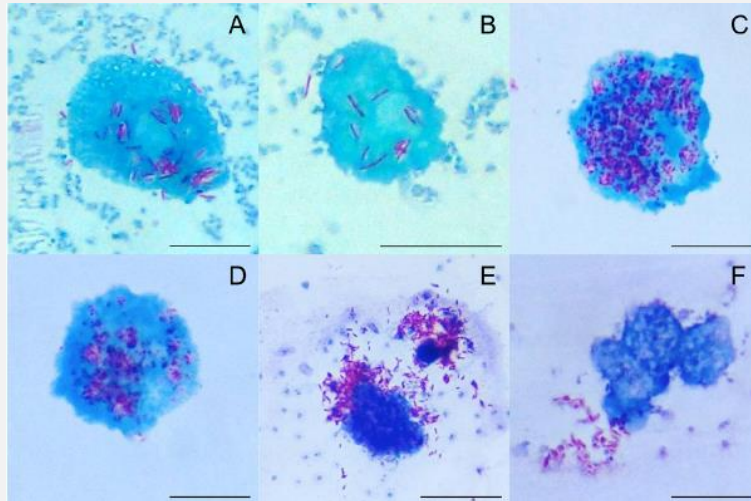
WHAT'S IN YOUR PIPES?



- organisms are many times more resistant to chlorine, chloramine, chlorine dioxide and ozone than other water borne micro-organisms such as *E. coli*.
- disinfection may contribute in part to the persistence of MAC, as competition for growth is lacking.
- significantly higher mycobacterial numbers in distribution samples (average 25000 fold) than those collected immediately downstream from treatment plant

NTM SURVIVE WITHIN AMOEBAE

- vehicle for replication and transmission
- enhanced virulence

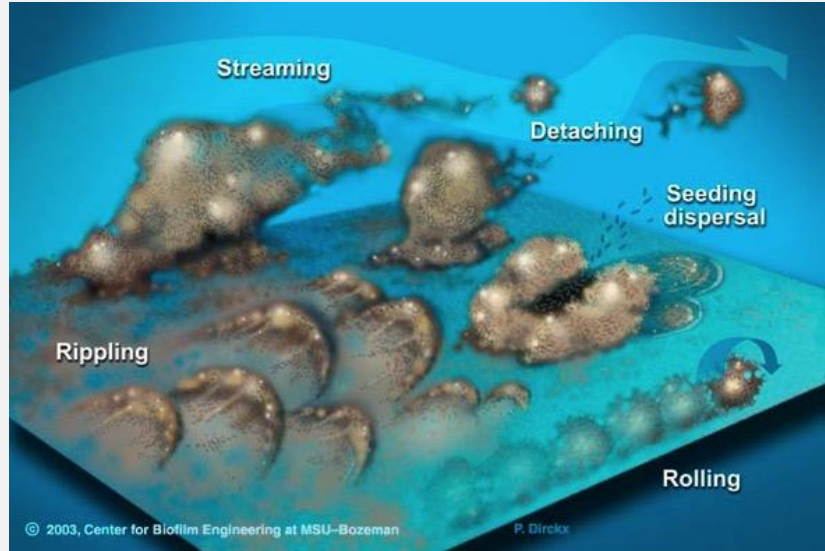


A,B *Acanthamoeba* spp
C,D *Protacanthamoeba bohemica*
E,F *Vermamoeba vermiformis*

Delafont Environ Sci Technol 2014

RELEASE FROM BIOFILMS

- **Transient pressure events**
- Rapid changes in water flow rate – increase the concentrations of bacteria in water.
- This occurs when we turn taps on and off
- Greater in smaller diameter pipes



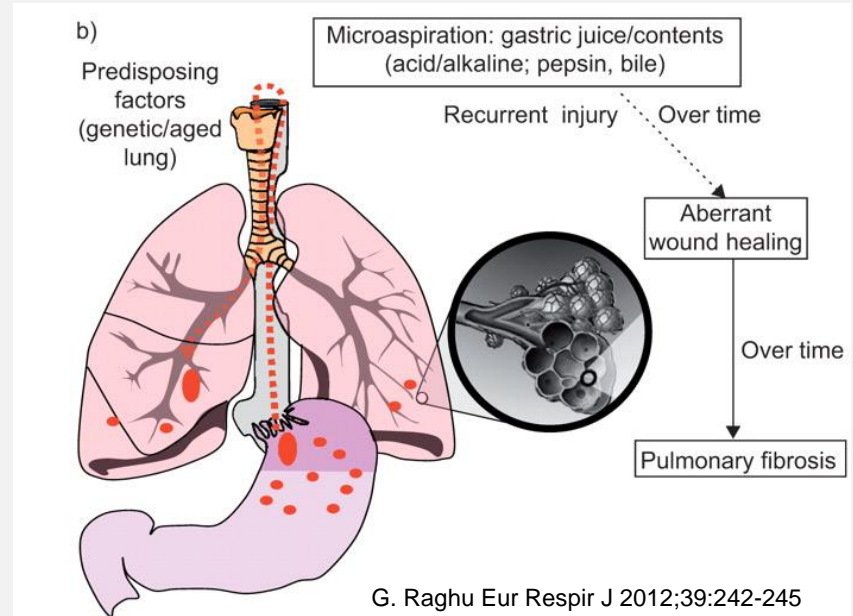
POINT OF USE CONNECTIONS

- POU connections and treatment devices promote biofilm formation, and amplify numbers of mycobacteria.



ASPIRATION OR INHALATION ?

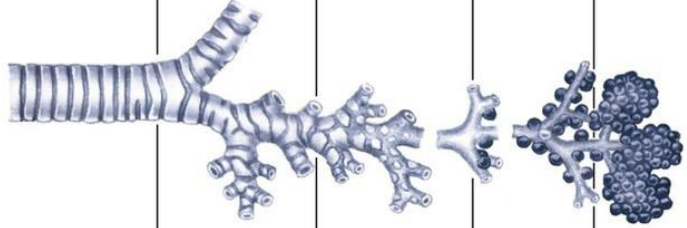
- Several studies associating NTM disease with GORD
- Deposition of relatively few mycobacteria can infect the host –2 to 3 bacteria, up to 50
- Aspiration of upper respiratory tract bacteria into the lungs -~ 50% of normal hosts during sleep
- 15.5% patients with MAC aspiration was a clinical problem

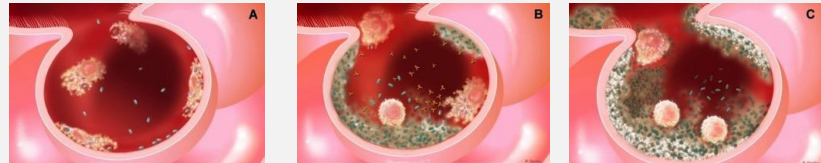


AEROSOLS



- Mycobacteria can be aerosolized from natural waters and transferred from seawater to air by natural processes.
- **Aerosolisation- 1 000 fold increase** in the numbers of viable mycobacterial cells per ml of water
- Mycobacteria in natural aerosols found in particles and **droplets of a respirable size (ie <math><5\mu\text{m}</math>)**

Conducting Airways			Respiratory Unit	
Trachea	Segmental bronchi	Subsegmental bronchi (bronchioles)		Alveolar ducts
		Nonrespiratory	Respiratory	
				
Generations	8	16	24	26



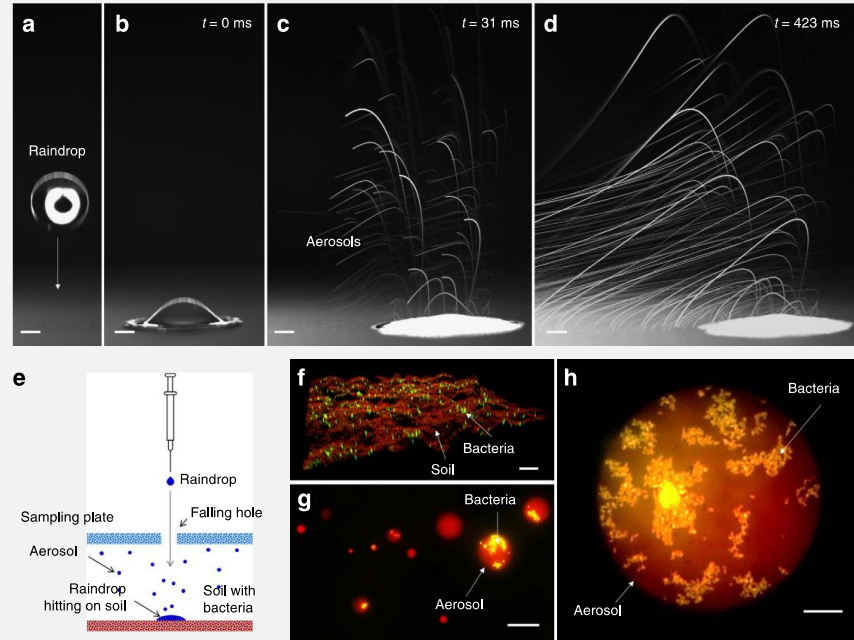
AEROSOLS



- Hypersensitivity pneumonitis – use of hot tubs, ***M. avium***
- reported in lifeguards and pool attendants, who worked in an indoor swimming pool that featured waterfalls and sprays.
- following disinfection of metalworking fluid ***M. immunogenum***

BIOAEROSOL GENERATION BY RAINDROPS ON SOIL

- bioaerosols -aerosols containing microbes
- after a rainfall - rapid increase of bioaerosol concentration in the air
- hundreds of aerosols are generated from a single raindrop within a few microseconds
- estimate number of bacteria that can be transferred by aerosol as a function of soil type, bacterial surface density, rainfall velocity and soil surface temperature.



Bioaerosols transported by wind to other places much faster than other modes of transfer

SHOWERHEADS

- Culture-independent technology (rRNA gene sequences)
- Sequences representative of **nontuberculous mycobacteria (NTM)** and other opportunistic human pathogens are **enriched to high levels in many showerhead biofilms**, >100-fold above background water contents.

Feazel *PNAS* 2009, *106*, 16393-16399.

Journal of
Clinical Microbiology

Isolation of Nontuberculous Mycobacteria (NTM) from Household Water and Shower Aerosols in Patients with Pulmonary Disease Caused by NTM

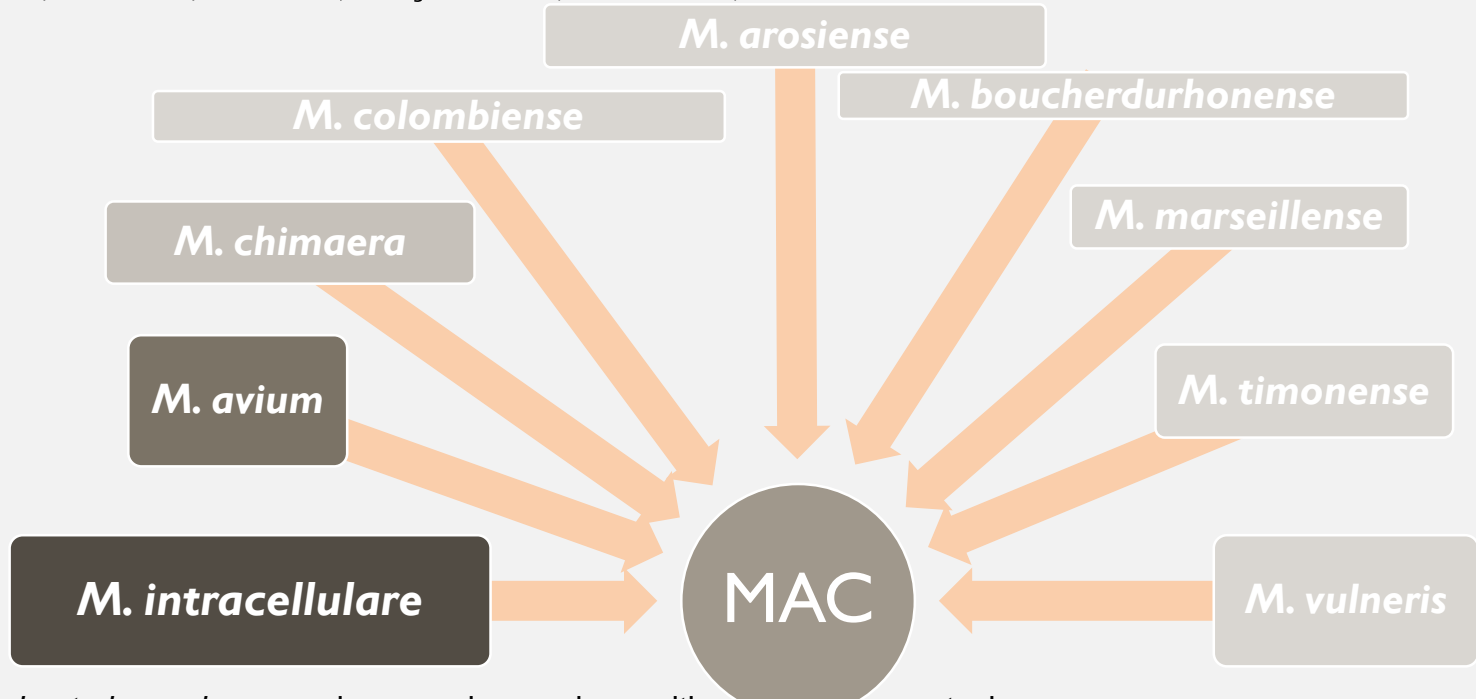
Rachel Thomson, Carla Tolson, Robyn Carter, Chris Coulter, Flavia Huygens and Megan Hargreaves
J. Clin. Microbiol. 2013, *51*(9):3006. DOI:
10.1128/JCM.00899-13.
Published Ahead of Print 10 July 2013.



- NTM grown from shower aerosols: *M. avium*, *M. abscessus*, *M. kansasii*

Absence of *Mycobacterium intracellulare* and Presence of *Mycobacterium chimaera* in Household Water and Biofilm Samples of Patients in the United States with *Mycobacterium avium* Complex Respiratory Disease

Richard J. Wallace, Jr.,^a Elena Iakhiaeva,^a Myra D. Williams,^b Barbara A. Brown-Elliott,^a Sruthi Vasireddy,^a Ravikiran Vasireddy,^a Leah Lande,^{c,d} Donald D. Peterson,^{c,d} Janet Sawicki,^d Rebecca Kwait,^c Wellington S. Tichenor,^e Christine Turenne,^f Joseph O. Falkinham III^b



Genotyping of *Mycobacterium avium* complex organisms using multispacer sequence typing

Caroline Cayrou, Christine Turenne, Marcel A. Behr and Michel Drancourt

Microbiology (2010), 156, 687–694

GENOTYPIC MATCHING OF CLINICAL AND ENVIRONMENTAL ISOLATES

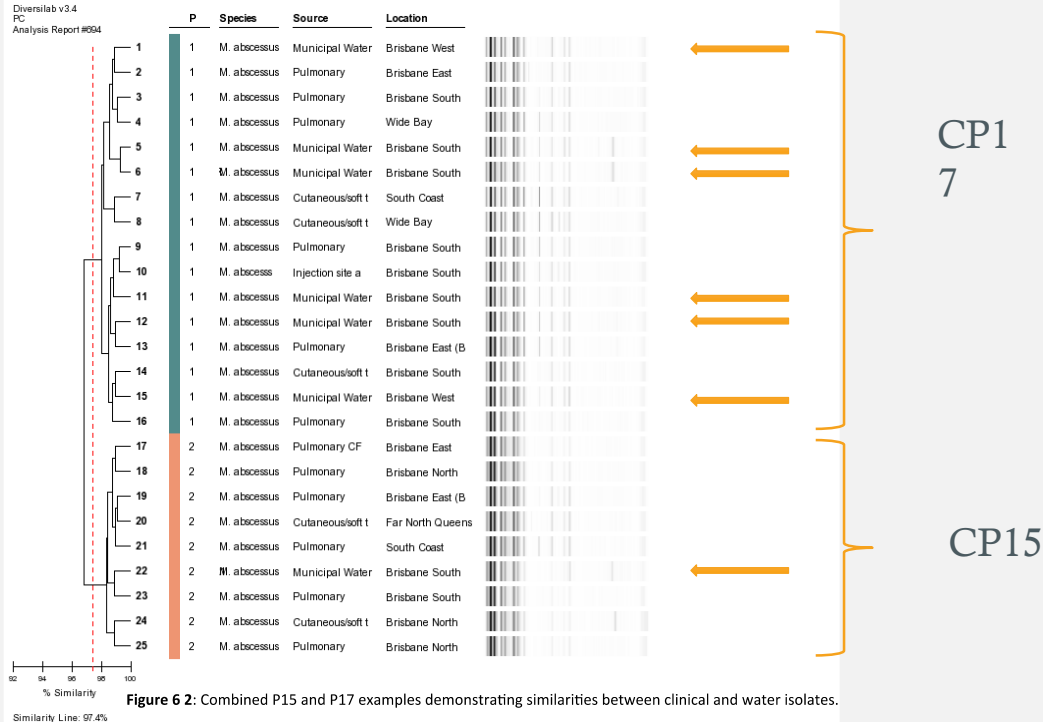


Figure 6 2: Combined P15 and P17 examples demonstrating similarities between clinical and water isolates.

SOIL

TABLE 2.

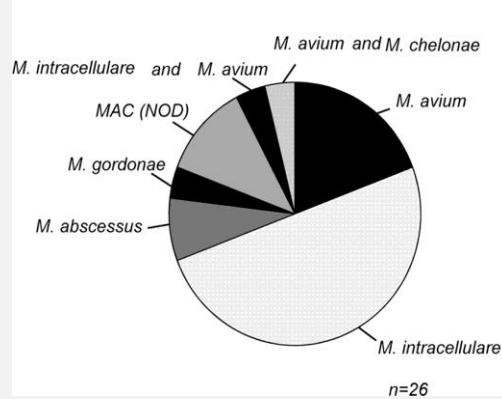
Mycobacterial species isolated from pulmonary MAC patients

Species isolated from soil	<i>M. avium</i> (n = 67)	<i>M. intracellulare</i> (n = 26)	<i>M. avium</i> + <i>M. intracellulare</i> (n = 7)	p value
MAC	36 (53.7) ^a	10 (38.5) ^a	4 (57.1)	0.38
<i>M. avium</i>	23 (34.3)	4 (15.4)	1 (14.3)	0.19
<i>M. intracellulare</i>	17 (25.3)	7 (26.9)	3 (42.9)	0.36

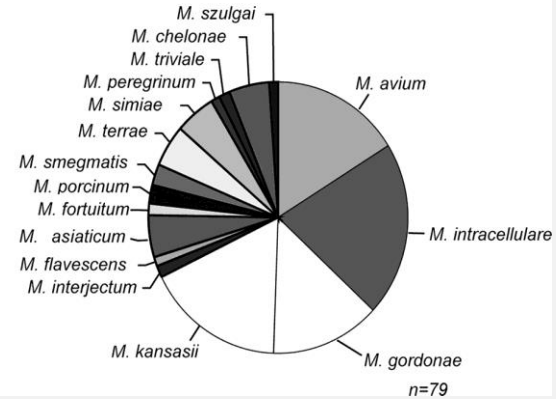
Data show number (%) of samples positive for MAC strains.

^aBoth *M. avium* and *M. intracellulare* were detected in four soil samples from the residences of patients infected with *M. avium* and in one soil sample from the residence of a patient infected with *M. intracellulare*.

A. Patient isolates



B. Soil isolates





NTM grown from aerosols generated by pouring of potting mix and soil samples from patient homes (not speciated)

At least 3 studies that have correlated soil exposure with NTM risk

ENVIRONMENTAL RISK FACTORS FOR INFECTION WITH MYCOBACTERIUM AVIUM COMPLEX

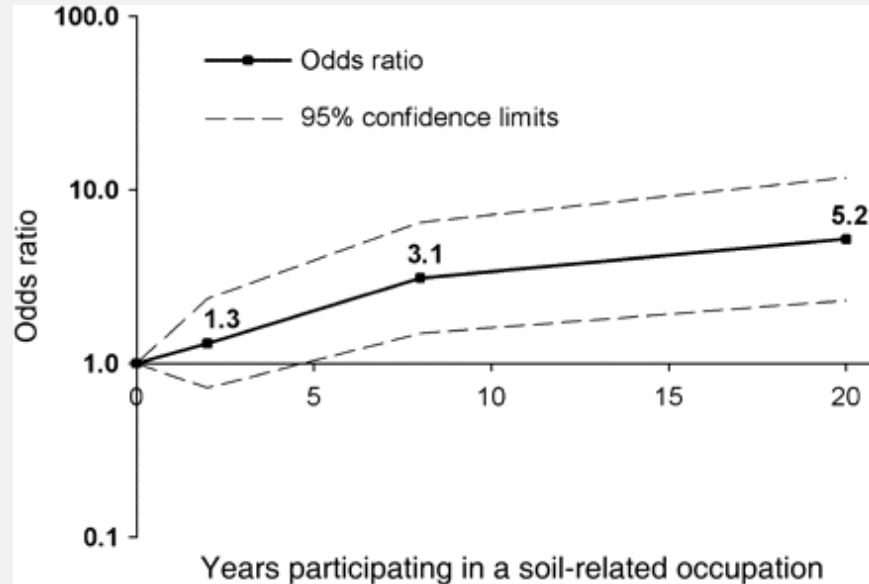
Soil exposure

Soil occupation, any	94/147	83/186	53.1	2.2	1.4, 3.4	<0.01
None	53/147	103/186	34.0	1.0		
Farm planting, ever‡	55/146	41/185	57.3	2.6	1.5, 4.4	<0.01
Farm truck driver, ever†,‡	33/139	25/172	56.9	2.6	1.4, 4.8	<0.01
Lawn/landscape service, ever‡	34/146	28/186	54.8	2.4	1.3, 4.3	<0.01
Other soil occupations, ever‡	33/145	26/185	55.9	2.5	1.3, 4.5	<0.01

Water exposures

Shower/bath >once/day	59/147	63/186	48.4	1.3	0.8, 2.0	0.24
Does dishes by hand >once/day	42/123	56/145	42.9	0.8	0.5, 1.4	0.45
Drinks bottled water >once/day	28/136	38/163	42.4	0.9	0.5, 1.5	0.57
Ever swims, any	107/147	145/186	42.5	0.8	0.5, 1.6	0.28
Ever, pool	73/141	109/183	40.1	0.7	0.5, 1.1	0.16
Ever, lake	31/138	48/179	39.2	0.8	0.5, 1.6	0.37
Ever, ocean	91/144	119/181	43.3	0.9	0.6, 1.4	0.63

ENVIRONMENTAL RISK FACTORS FOR INFECTION WITH *MYCOBACTERIUM AVIUM* COMPLEX



Reed C Am. J. Epidemiol. **2006**, 164, 32-40.

HOUSE-DUST

Dust particles are commonly suspended in the air

120 strains NTM isolated from vacuum cleaner dust collected in SE QLD

50 strains of *M. intracellulare* – 44% belonged to serotypes that were recognized as disease associated strains

Dawson MJA 1971

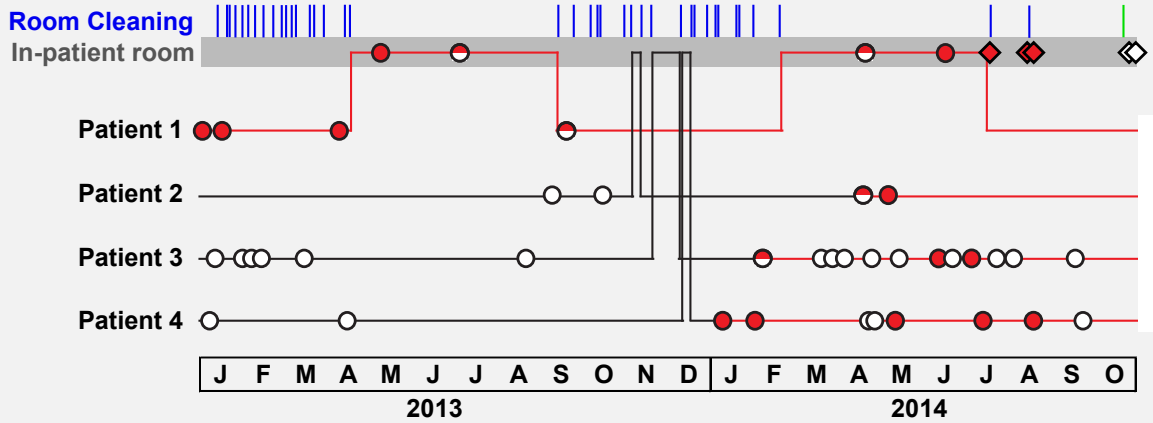


NTM IN HOUSE DUST

Mycobacterial species isolated from the dust samples. Combined data of the GLC and 16 S rDNA sequence analyses.

Species or groups	N	%
<i>M. terrae</i> complex (<i>M. nonchromogenicum</i> or <i>M. arupense</i> , <i>M. kumamotoense</i> , and <i>M. terrae</i>)	43	31
<i>M. avium. intracellulare, scrofulaceum</i> complex (<i>M. colombiense</i> , <i>M. intracellulare</i> , <i>M. avium</i> , and <i>M. scrofulaceum</i>)	38	27
<i>M. gordonae</i>	22	16
<i>M. lentiflavum</i>	7	5
<i>M. triviale</i>	5	4
<i>M. asiaticum</i>	3	2
<i>M. "aquaeductus"</i>	3	2
<i>M. bohemicum</i>	2	1
<i>M. interjectum</i>	2	1
<i>M. malmoense</i>	2	1
<i>M. palustre</i>	1	1
<i>M. "savoniense"</i>	1	1
<i>M. tusciae</i>	1	1
Non-identifiable	10	7

FOMITE SPREAD

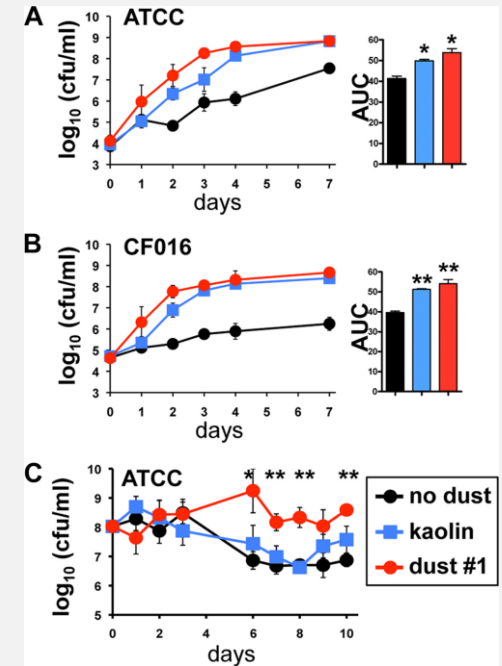


- | | | |
|------------------------------------|------------------------------|--------------------------|
| Sputum samples | Environmental samples | Room Cleaning |
| ○ Smear negative, culture negative | ◇ Culture negative | Standard regimen |
| ◐ Smear negative, culture positive | ◐ Culture positive | Clinell™/perchlorate/HPV |
| ● Smear positive, culture positive | | |



FOMITES – NON LIVING CARRIERS OF INFECTIOUS AGENTS

- *M. abscessus* displays fitness for fomite transmission
- *M. abscessus* physically associated with particulates
- Growth enhanced in the presence of both kaolin and house dust
- Survived desiccation for 2 weeks, but not viable after 3 weeks
- Rate of decline of *M. abscessus* viability during desiccation was reduced in the presence of house dust
- Evidence for enhanced growth and survival of *M. abscessus* during alternating growth and drying periods



REDUCING EXPOSURE – WHY?

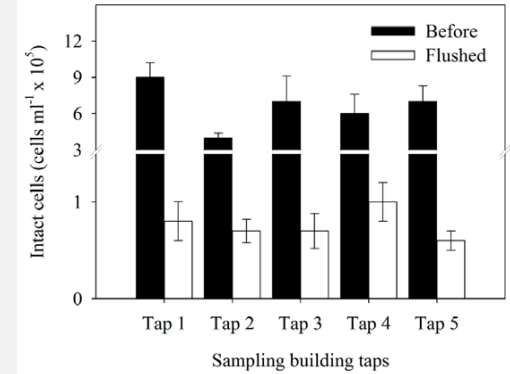
- Prevent new infections in susceptible patients
- Prevent re-infection whilst on treatment and after completion of treatment
- Recurrence rates as high as 30-48% after apparently successful treatment
- Most recurrences are new infections with a different strain type of the same species or a different species
- Many patients with nodular bronchiectasis can have different strains present in the airways at the same time, that appear to come and go over time

EVIDENCE

- The exact route of acquisition in the individual patient – e.g dust v shower aerosol v soil v aspiration
- How much exposure is necessary- i.e. what dose of organism is needed?
- Single vs repetitive exposures
- Impact of climate and industry on background environmental levels of NTM
- No formal studies of any intervention to reduce NTM exposure
- These studies are difficult to conduct and common sense should prevail
- Any intervention that is simple, cost effective and easy to implement and that has some evidence base in theory should be considered

FLUSHING OF TAPS

- Stagnation of water in pipes leading to taps that are infrequently used has been shown to increase NTM in 'first catch' samples
- Can simply be overnight stagnation
- For taps used infrequently, a **5 minute flush** is recommended before use of the water from that tap
- For taps used on a daily basis, a **shorter flush on first use for the day** is recommended (e.g. 200ml)



REMOVAL OF MESH AERATORS

- Mycobacteria attach to the large surface area provided by the mesh, form biofilm in which they can then proliferate
- Numbers of mycobacteria coming through taps with them attached are greater than taps without
- Their removal may help to reduce exposure from water

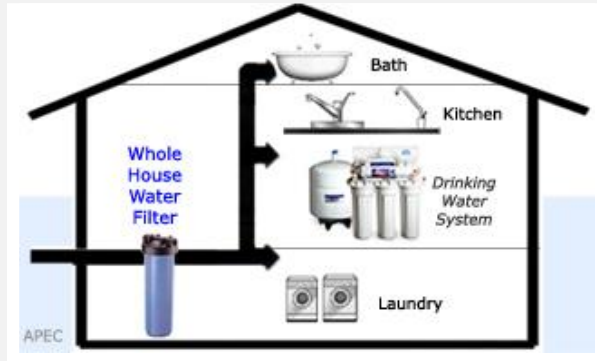


POINT OF USE FILTERS

- NTM can multiply within carbon filters, even those impregnated with silver
- NTM are released into the water passing through
- Avoid
- If they are to be used, they need to be changed every 2 weeks



UNDERBENCH/PRE-TAP WATER FILTRATION

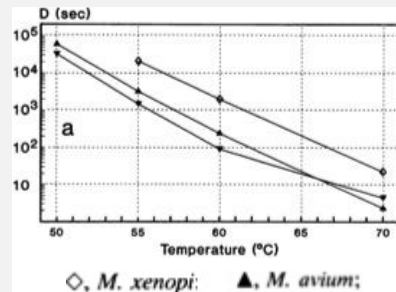


- Cartridge based systems installed under kitchen benches or anywhere after the main supply of water enters the premises
- Microfiltered water dispensers (MWDs) with composite filters
- Reverse-osmosis water dispensers (ROWDs)
- Adequate and continuous maintenance is crucial
- May filter other bacteria, chemicals and sediment
- Have been shown to increase the numbers of mycobacteria in tap water



TURN UP THE HEAT

- Hot water should be stored at 60°C (140°F) minimum
- many can heat up to 75-80°C
- A tempering device or thermostatic mixing valve can be installed at the tap to ensure the maximum temperature of water deliver from the tap is 50°C to reduce scalding risk (45° is recommended for children and the elderly)

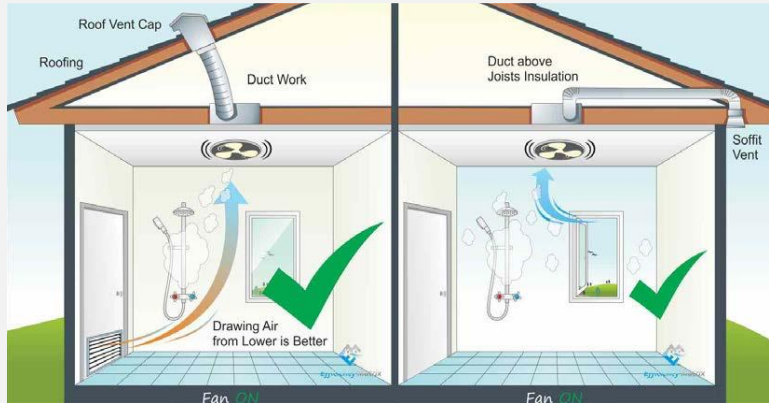


SHOWERS



- Replace plastic showerheads with metal ones (less likely to harbor biofilm where NTM grow)
- Soaking shower heads in vinegar for 60 minutes has been recommended as a means of cleaning
- Needs to be repeated at least every 6 months (probably more often)
- Household bleach has been recommended as an alternative - however suggestion of regrowth of NTM with increased resistance to chlorine

VENTILATION



- Ensure adequate bathroom ventilation to clear contaminated shower aerosols and steam
- The use of exhaust fans, and leaving windows open with help alleviate steam build up
- Replace/clean filters on exhaust fans as per manufacturers instructions (usually every 6 mths)

NEBULIZERS AND AIRWAY CLEARANCE DEVICES

- Nebulizers should be cleaned after each use
- The use of unit dose vials for medication is recommended (rather than a larger volume that is repeatedly accessed, when contamination can occur)
- Heat methods:
 - Steam sterilizer marketed for baby bottles
 - Place in boiling water and boil for 5 minutes
 - Place in microwave- safe container submerged in water and microwave for 5 min
 - Use dishwasher if temp $>70^{\circ}\text{C}$ (158°F) for 30 min



OTHER DEVICES

- CPAP humidifiers, neti pots/nasal rinse devices, humidifiers and other devices that require water for humidification/aerosolization, should be used with sterile/boiled water only



OTHER WATER RELATED EXPOSURES

- Indoor pools, hot tubs, steam saunas, hydrotherapy pools and other leisure activities that involve aerosolization of water should be avoided



REFLUX AND NOCTURNAL MICRO-ASPIRATION

- Maintain healthy weight, avoid tight fitting clothing
- Avoid foods and drinks that trigger heartburn
- Eat smaller meals more frequently (to maintain caloric requirements)
- Don't lie down after a meal. Wait at least 3 hours after eating before lying down or going to bed
- Use a foam wedge to elevate the head of the bed



SOIL

- No studies examining the effectiveness of masks
- Reasonable for high risk patients to use personal-protection whilst engaging in dusty activities
- Moistening soil/potting mix prior to use may reduce the aerosolization and subsequent inhalation of contaminated soil particles



SUMMARY

- NTM causing human infections can be found in a wide variety of environmental sources, but environmental niche can be species specific – eg *M. avium* vs *M. intracellulare*
- Geographic variation in species distribution in both patients and the environment
- Human activities may influence the numbers of mycobacteria in the environment (e.g agriculture, water disinfection practices) and increase exposure (e.g hot tub use, soil related activities, house dust)
- Aerosols of a respirable particle size generated by showering and the pouring of potting mix/soil can be inhaled
- *M. intracellulare* – the main pathogen responsible for NTM disease in most parts of the world is not found in water/showerheads
- No evidence that showering frequency is associated with NTM infection
- Climate and natural events likely to influence mycobacterial numbers in the environment and more work is need in this space



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







Luke Knibbs
Archie Clements
Michael Chou



Andres Floto
Dorothy Grogono
Nicole Wheeler

SURVIVAL OF NTM ON SURFACES

- MTB can survive on dry inanimate surfaces 1 day- 4mths

Levels of Disinfection Difficulty		
<div style="writing-mode: vertical-rl; transform: rotate(180deg);">Harder to Kill</div>	<div style="writing-mode: vertical-rl; transform: rotate(180deg);">Extremely Hard to kill</div>	Prions  Transmissible Spongiform Encephalopathy (TSE); Creutzfeldt-Jakob disease(CJD) Mad cow disease; Scrapies
	Bacterial Spores  Spores of: C. difficile; C. tetanus; C. botulinum; C. perfringens; Anthrax	
	Mycobacteria  M. tuberculosis; M. avium	
	Viruses without envelopes  Norovirus; Rotavirus; Rhinovirus; Poliovirus; Papillomavirus (HPV); Coxsackie; Adenovirus	
	Fungi includes fungal spores  Aspergillus fumigatus, A. flavus; A. niger; Candida albicans	
	Gram negative bacteria  Pseudomonas; Acinetobacter, Klebsiella, E. coli; Enterobacteriaceae, Legionella	
	Gram positive bacteria  Staphylococcus; Enterococcus; Streptococcus; Clostridia vegetative rods	
	Viruses with lipid envelopes  Influenza; HBV; HCV; HIV; RSV; Coronavirus; CMV; HSV; Measles; Mumps; Rubella; VZV (Varicella-Zoster) Shingles/ Chickenpox	
	<div style="writing-mode: vertical-rl; transform: rotate(180deg);">Easy to kill</div>	

